Total Synthesis of the Cytotoxic *Threo, Trans, Threo, Trans, Threo* Annonaceous Acetogenin Asimin and Its C-10 Epimer: Unambiguous Confirmation of Absolute Stereochemistry

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A convergent synthesis of asimin (1) and its C-10 epimer **33** is reported. The essential features of this synthesis include (a) the addition of an enantioenriched γ -OMOM allylic indium reagent to a core C-23 aldehyde precursor to install the C-24–C-34 segment with concomitant introduction of the C-24 and C-23 stereocenters; (b) the addition of an enantioenriched γ -OMOM allylic indium reagent to a core C-16 aldehyde to install the C-10–C-15 segment with formation of the C-15 and C-16 stereocenters, (c) the addition of a dialkyl zinc reagent, catalyzed by a chiral triflamide-Ti(O-*i*-Pr)₄ complex, to introduce the C-1–C-9 segment with creation of either the 10(*R*) or 10(*S*) stereocenters; and (d) aldol condensation of the foregoing C-1–C-34 segment with OTBS-protected lactic aldehyde to incorporate the C-35–C-37 butenolide segment. Removal of the three MOM protecting groups was achieved with aqueous HCl in THF. The 10(*R*) diastereomer was found to correspond to natural asimin.

The Annonaceous acetogenins comprise a widespread family of cytotoxic natural products with a remarkable range of biological activities.¹ Their novel and selective mode of action as inhibitors of oxidative phosphorylation offers a unique potential for these compounds as anticancer agents.² In fact, data from cell-culture bioassays against human tumor cell lines reveals nearly unbelievable levels of cytotoxicity, even against cells exhibiting multiple-drug resistance toward current chemotherapeutic agents.³ For this reason, and by virtue of their extremely limited availability, these compounds have been targeted for total synthesis by a number of research groups.⁴

In 1994, McLaughlin and co-workers reported the isolation of three acetogenins, asimin (1), asiminacin (2), and asiminecin (3), from the stem back of the North American paw-paw tree, Asimina triloba Dunal,⁵ which they identified as structural isomers of asimicin (4), a compound previously isolated from the same source.⁶ All three of the new compounds showed extremely high potency against human tumor cell lines. The initial report depicted asimin as the C-10(*S*) isomer. However, in a subsequent paper the stereochemistry at C-10 was shown as *R*.⁷ In that report, it was revealed that the assignment of configuration was based upon chemical shift differences of 0.06 Hz between the protons at C-4 and C-3 in the ¹H NMR spectra of the tris(2-methoxy-2-trifluoromethyl-2-phenylacetate) (MTPA) derivatives.⁸ In view of the rather subtle basis for this assignment, and because of the high biological profile of these compounds, we undertook a total synthesis of both C-10 epimers of asimin along lines previously developed in our laboratory.^{4c,9} Our synthesis started with alcohol **5**, which was converted to aldehyde 6 as previously described.¹⁰ Addition of an allenylindium intermediate, generated in situ from the (*R*)-allylic stannane 7 and InCl₃, to aldehyde 6 of >95% ee afforded alcohol 8 (Scheme 1).4c This alcohol was "protected" as the tosylate derivative 9. Upon stirring with 5% Pd-C under an atmosphere of hydrogen, tosylate 9 underwent hydrogenation of the double bond and concomitant hydrogenolysis of the benzyl ether. The resulting alcohol **10** was oxidized with PCC¹¹ to afford aldehyde 11 in high overall yield. Addition of the allylic indium reagent prepared by treatment of the (*S*)-allylic stannane **12** in situ with $InCl_3$ yielded the adduct **13**.^{4c} The bistosylate **14** gave rise to the *threo*, *trans*, *threo*, *trans*, *threo* bis-tetrahydrofuran core unit **15** upon stirring with TBAF in THF. Hydrogenation/hydrogenolysis with H₂/Pd-C at one atmosphere afforded alcohol **16**, which was oxidized to the aldehyde **17** with PCC.¹¹

The C-1-C-9 side chain of asimin was introduced through addition of the organozinc reagent 18 to aldehyde **17** in the presence of $Ti(O-i-Pr)_4$ and the (S,S)-cyclohexanediamine bis-triflamide catalyst **19** (Scheme 2).^{4c,12} The resulting adduct, alcohol 20 is presumed to be the 10Rdiastereomer based on a wealth of precedent.¹² Attempted assignment of C-10 absolute configuration through analysis of the ¹H NMR spectra of the (R)- and (S)-O-methyl mandelates was not successful owing to the absence of diagnostic peaks.¹³ The MOM ether **21** was condensed with the TBS ether of (S)-lactic aldehyde 22 to afford, after treatment with TBAF, the γ -lactone adduct **23** as a mixture of C35 isomers.^{4e,14} Exposure of the alcohol **23** to trifluoroacetic anhydride and triethylamine led to the butenolide 25 via the unisolated trifluoroacetate 24. Cleavage of the MOM ethers was effected with aqueous HCl in THF. For comparison purposes, the triol **1** was converted to the (*S*)-MTPA ester 26.8 The ¹H NMR spectra of triol 1 and the (S)-ester 26 were identical to the corresponding spectra of asimin and its (S)-MTPA ester. The authentic (R)-MTPA ester showed several small differences in the ¹H NMR spectrum. In addition, the optical rotation of the triol 1 was in good agreement with the reported value.⁵

10(*S*)-Asimin (**33**) was prepared from aldehyde **17** by an identical sequence, except the triflamide **27** of (*R*,*R*)-1,2-cyclohexanediamine was used to catalyze the addition of the alkylzinc reagent **18** to aldehyde **17** (Scheme 3). The (*S*)-MTPA derivative **34** was subtly, but definitely different from the (*S*)-MTPA derivative **26** of the 10*R* alcohol **1**. In addition, the optical rotation was significantly lower for the 10*S* compound ($[\alpha]_D$ +22 for **1** and +16 for **33**). Interestingly, the ¹H NMR spectrum of the (*S*)-MTPA derivative **34** was virtually identical to that of the (*R*)-MTPA derivative **34** was virtually identical to that of the (*R*)-MTPA derivative **34** was virtually identical to that of the (*R*)-MTPA derivative for the 10*R* alcohol, indicating a pseudoenantiomeric relationship between these two derivatives. This finding



Asimin (1) $R^1 = R^2 = R^4 = H$; $R^3 = (R)$ -OH Asiminacin (2) $R^1 = R^3 = R^4 = H$; $R^2 = OH$ Asiminecin (3) $R^1 = OH$; $R^2 = R^3 = R^4 = H$ Asimicin (4) $R^1 = R^2 = R^3 = H$; $R^4 = OH$

Figure 1. Representative members of the asimicin subgroup of Annonaceous acetogenins.

Scheme 1



a) (MeO)₃CMe, EtCO₂H, heat (96%); b) AD-mix β (99%);c) MeONHMe•HCl, AlMe₃ (99%); d) TBSCl, Im (99%); e) DIBAL-H (99%)

Scheme 2



Scheme 3



is not surprising considering the ¹H NMR spectra of the diastereomeric alcohols **1** and **33** are identical.

On the basis of these results, the structure assigned to asimin (1) by McLaughlin and co-workers through analysis of the MTPA ester ¹H NMR chemical shift differences can be taken as correct.⁷ A noteworthy feature of the present synthesis is the ability to "protect" the alcohol functions of adduct **8** as the tosylate **9** for later use in the bis-cyclization reaction $14 \rightarrow 15$.

Experimental Section

General Experimental Procedures. NMR spectra were determined in CDCl₃ at 300 MHz (¹H NMR) and 75 MHz (¹³C NMR). The chemical shifts are expressed in δ values relative to TMS. FT–IR were taken on an Impact 410 spectrometer. Optical rotations were measured on a Perkin–Elmer 343 polarimeter. Kieselgel 60F₂₅₄ plates were employed for TLC analyses. Si gel (200–400 mesh) was used for column chromatography. Reagents prepared according to literature procedures are footnoted. All other reagents were obtained from commercial sources. All reactions were performed under N₂ in oven-dried flasks. Elemental analyses were carried out by Atlantic Microlab, Inc. (Norcross, GA).

Alcohol 8. A solution of 1.07 g (4.84 mmol) of InCl₃ in 80.0 mL of EtOAc was sonicated for 20 min, and to it was added a solution of 2.10 g (4.24 mmol) of aldehyde 64c in 2.0 mL of EtOAc. The mixture was cooled to -78 °C, and to it was added a solution of 3.43 g (6.81 mmol) of stannane $7^{\rm 4c}$ in 2.0 mL of EtOAc. The reaction mixture was allowed to warm to room temperature (3 h), quenched with NaHCO₃, and extracted with ether. The ether extracts were washed with brine, dried with MgSO₄, and concentrated under reduced pressure. The crude product was purified by column chromatography on Si gel (elution with 10% EtOAc in hexane) to afford 2.50 g (83%) of alcohol 8: $[\alpha]^{25}_{D}$ -9.7 (c 0.66, CHCl₃); IR (film) 3484, 2916 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 7.33 (5 H, m), 5.72 (1 H, dt, J = 6.9, 15.8 Hz), 5.38 (1 H, dd, J = 9.1, 15.8 Hz), 4.73 (1 H, d, J = 7.3 Hz), 4.56 (1 H, d, J = 7.3 Hz), 4.50 (2 H, s), 3.92 (1 H, m), 3.65 (1 H, m), 3.56 (2 H, m), 3.47 (2 H, t, J = 6.6 Hz), 3.37 (3 H, s), 2.06 (2 H, m), 1.94-1.12 (20 H, m), 0.88 (21 H, m), 0.05 (6 H, s), 0.04 (6 H, s); 13 C NMR (CDCl₃, 75 MHz) δ 138.7, 137.5, 128.3, 127.5, 127.4, 124.8, 93.6, 80.4, 75.8, 75.4, 74.2, 72.7, 70.7, 55.4, 32.4, 31.9, 29.9, 29.4, 29.2, 29.1, 27.1, 26.7, 26.6, 25.8, 22.6, 18.0, 14.1, -4.1, -4.6; anal. C 67.57%, H 10.72%, calcd for C40H76O6Si2, C 67.74%, H 10.80%.

Tosylate 9. To a mixture of 1.00 g (1.41 mmol) of alcohol 8 in 2.0 mL of pyridine was added 1.60 g (8.4 mmol) of p-TsCl. The reaction mixture was stirred for $1\tilde{2}$ h, quenched with H_2O and extracted with ether. The ether extracts were washed with brine, dried over MgSO₄, and concentrated under reduced pressure. The crude product was purified by column chromatography on Si gel (elution with 10% EtOAc in hexane) to afford 1.14 g (94%) of tosylate **9**: $[\alpha]^{25}_{D}$ –13.4 (*c* 0.50, CHCl₃); ¹H NMR (CDCl₃, 300 MHz) δ 7.78 (2 H, d, J = 8.5 Hz), 7.30 (7 H, m), 5.70 (1 H, dt, J = 6.1, 15.2 Hz), 5.20 (1 H, dd, J = 7.6, 15.4 Hz), 4.61 (1 H, d, J = 6.5 Hz), 4.50 (2 H, s), 4.45 (1 H, d, J = 6.5 Hz), 4.44 (1 H, m), 4.25 (1 H, m), 3.43 (4 H, m), 3.32 (3 H, s), 2.38 (3 H, s), 2.00 (2 H, m), 1.87-1.15 (20 H, m), 0.85 (21 H, m), 0.03 (6 H, s), 0.00 (6 H, s); ¹³C NMR (CDCl₃, 75 MHz) & 144.1, 138.6, 137.3, 134.5, 129.5, 128.3, 127.9, 127.6, 127.4, 124.7, 93.6, 86.0, 77.6, 75.4, 75.0, 72.8, 70.6, 55.4, 32.3, 31.8, 29.4, 29.2, 28.9, 27.0, 26.8, 26.5, 26.4, 26.0, 25.8, 25.7, 22.6, 21.5, 17.9, 14.1, -4.1, -4.6.

Alcohol 10. A mixture of 0.40 g (0.46 mmol) of tosylate **9** and 0.40 g of Pd-C (5%) in 6.0 mL of EtOAc was placed under one atmosphere of H₂ (balloon). The reaction mixture was stirred for 12 h and filtered through Celite. Solvent was removed under reduced pressure, and the crude product was purified by column chromatography on Si gel (elution with 30% EtOAc in hexane) to afford 0.34 g (94%) of alcohol **10**: $[\alpha]^{25}_{D}$ +22.0 (*c* 0.50, CHCl₃); IR (film) 3493 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 7.78 (2 H, d, *J* = 8.5 Hz), 7.31 (2 H, d, *J* = 8.5 Hz), 4.68 (1 H, d, *J* = 6.5 Hz), 4.58 (1 H, d, *J* = 6.5 Hz), 4.46

(1 H, m), 3.80 (1 H, m), 3.63 (2 H, m), 3.49 (1 H, m), 3.40 (1 H, m), 3.35 (3 H, s), 2.43 (3 H, s), 1.80–1.15 (26 H, m), 0.86 (21 H, m), 0.04 (6 H, s), 0.02 (6 H, s); 13 C NMR (CDCl₃, 75 MHz) δ 144.3, 134.3, 129.6, 127.9, 96.4, 85.8, 78.3, 77.6, 77.2, 76.6, 75.3, 75.0, 63.1, 55.7, 31.9, 31.4, 30.0, 29.6, 29.4, 29.3, 26.6, 26.3, 26.1, 25.8, 25.7, 25.6, 22.7, 21.6, 17.9, 16.9, 15.2, 14.1, -4.1, -4.2, -4.7.

Aldehyde 11. To a mixture of 0.55 g (0.71 mmol) of alcohol 10 and 0.40 g of 4 Å molecular sieves in 6.0 mL of CH₂Cl₂ at 0 °C was added 0.50 g (2.30 mmol) of PCC. The reaction mixture was stirred at room temperature for 1 h, diluted with ether, and filtered through Celite. Solvent was removed under reduced pressure, and the residue was purified by column chromatography on Si gel (elution with 10% EtOAc in hexane) to afford 0.48 g (89%) of aldehyde 11: $[\alpha]^{25}_{D}$ +22.0 (c 0.62, CHCl₃); IR (film) 1728 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 9.77 (1 H, s), 7.79 (2 H, d, J = 8.5 Hz), 7.31 (2 H, d, J = 8.5 Hz), 4.68 (1 H, d, J = 6.5 Hz), 4.59 (1 H, d, J = 6.5 Hz), 4.47 (1 H, m), 3.81 (1 H, m), 3.49 (1 H, m), 3.42 (1 H, m), 3.36 (3 H, s), 2.45 (2 H, m), 2.42 (3 H, s), 1.91-1.13 (24 H, m), 0.87 (21 H, m), 0.03 (6 H, s), 0.02 (6 H, s); 13 C NMR (CDCl₃, 75 MHz) δ 202.3, 144.4, 134.3, 129.7, 127.9, 96.4, 85.7, 78.4, 75.2, 74.2, 55.7, 41.1, 31.9, 31.5, 29.6, 29.4, 29.3, 26.6, 26.0, 25.7, 22.6, 21.5, 17.9, 14.1, -4.1, -4.7, -4.8; anal. C 62.33%, H 10.03%, calcd for C40H76O8SSi2, C 62.13%, H 9.91%.

Alcohol 13. The procedure for alcohol 8 was employed with 0.17 g (0.77 mmol) of InCl₃, 0.59 g (0.76 mmol) of aldehyde 11, and 0.65 g (1.21 mmol) of stannane 12 in 8.0 mL of EtOAc. The crude product was purified by column chromatography on Si gel (elution with 15% EtOAc in hexane) to afford 0.67 g (86%) of alcohol 13: [α]²⁵_D -7.1 (*c* 0.76, CHCl₃); IR (film) 3510 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 7.78 (2 H, d, J = 8.5 Hz), 7.32 (7 H, m), 5.74 (1 H, dt, J = 6.5, 15.0 Hz), 5.41 (1 H, dd, J = 8.4, 15.4 Hz), 4.71 (1 H, d, J = 6.9 Hz), 4.67 (1 H, d, J = 6.9 Hz), 4.57 (1 H, d, J = 6.9 Hz), 4.55 (1 H, d, J = 6.9 Hz), 4.50 (2 H, s), 4.47 (3 H, m), 3.92 (1 H, m), 3.78 (1 H, m), 3.61 (1 H, m), 3.49 (3 H, m), 3.41 (1 H, m), 3.37 (3 H, s), 3.35 (3 H, s), 2.42 (3 H, s), 2.17 (2 H, m), 1.85-1.10 (28 H, m), 0.86(21 H, m), 0.02 (6 H, s), 0.00 (6 H, s); ¹³C NMR (CDCl₃, 75 MHz) δ 144.2, 139.5, 136.5, 134.4, 129.6, 128.3, 127.8, 127.5, 127.4, 125.7, 96.3, 93.7, 85.7, 80.5, 78.3, 75.5, 75.4, 74.0, 72.8, 69.6, 55.6, 55.4, 31.8, 31.4, 29.8, 29.5, 29.4, 29.2, 29.1, 26.6, 26.2, 25.7, 22.6, 21.5, 17.9, 14.0, -4.1, -4.2, -4.7, -4.8,

Bis-Tosylate 14. The procedure described for tosylate **9** was employed with 0.66 g (0.64 mmol) of alcohol 13, 0.73 g (3.8 mmol) of p-TsCl, and 1.5 mL of pyridine. The crude product was purified by column chromatography on Si gel (elution with 10% EtOAc in hexane) to afford 0.70 g (93%) of bis-tosylate 14: $[\alpha]^{25}_{D}$ –5.4 (*c* 0.47, CHCl₃); ¹H NMR (CDCl₃, 300 MHz) δ 7.79 (4 H, d, J = 8.5 Hz), 7.31 (9 H, m), 5.71 (1 H, dt, J = 6.5, 15.4 Hz), 5.23 (1 H, dd, J = 8.4, 15.5 Hz), 4.66 (1 H, d, J = 6.7 Hz), 4.57 (1 H, d, J = 6.7 Hz), 4.55 (1 H, d, J = 6.7 Hz), 4.53 (2 H, m), 4.49 (2 H, s), 4.40 (1 H, d, J = 6.7 Hz), 4.19 (1 H, m),3.75 (1 H, m), 3.47 (2 H, t, J = 6.1 Hz), 3.38 (2 H, m), 3.34 (3 H, s), 3.29 (3 H, s), 2.43 (3 H, s), 2.14 (2 H, m), 1.82-1.14 (28 H, m), 0.85 (21 H, m), 0.00 (12 H, m); ¹³C NMR (CDCl₃, 75 MHz) δ 144.2, 144.1, 138.5, 136.5, 134.6, 134.5, 129.6, 129.5, $128.3,\ 127.8,\ 127.5,\ 127.4,\ 125.2,\ 96.3,\ 93.5,\ 85.6,\ 85.4,\ 78.4,\\ 75.3,\ 75.2,\ 72.8,\ 69.5,\ 65.8,\ 55.7,\ 55.4,\ 31.9,\ 31.5,\ 29.6,\ 29.4,$ 29.3, 29.0, 28.9, 27.2, 26.6, 26.3, 25.7, 22.6, 21.6, 17.8, 15.2, 14.1, -4.1, -4.2, -4.7.

Bis-THF Olefin 15. To a solution of 1.10 g (0.93 mmol) of bis-tosylate **14** in 15.0 mL of THF was added 4.6 mL (4.6 mmol) of TBAF (1.0 M in THF). The reaction mixture was stirred at 50 °C for 12 h, quenched with H₂O, and extracted with ether. The ether extracts were washed with brine, dried over MgSO₄, and concentrated under reduced pressure. The crude product was purified by column chromatography on Si gel (elution with 20% EtOAc in hexane) to afford 0.42 g (75%) of bis-THF olefin **15**: $[\alpha]^{25}_{D}$ –13.3 (*c* 0.67, CHCl₃); IR (film) 2925, 2855, 1457 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 7.34 (5 H, m), 5.70 (1 H, dt, *J* = 6.9 Hz), 4.68 (1 H, d, *J* = 6.9 Hz), 4.58 (1 H, d, *J* = 6.9 Hz), 4.49 (2 H, s), 4.08–3.87 (6 H, m), 3.46 (2 H, t, *J* = 6.5 Hz), 3.39 (3 H, s),

3.37 (3 H, s), 2.15 (2 H, m), 2.00–1.15 (28 H, m), 0.88 (3 H, t, J = 7.2 Hz); ¹³C NMR (CDCl₃, 75 MHz) δ 138.4, 134.8, 128.2, 127.4, 127.0, 96.6, 93.5, 81.6, 81.3, 81.2, 81.0, 79.4, 78.5, 72.8, 69.5, 55.5, 55.1, 31.8, 37.0, 29.7, 29.5, 29.2, 29.1, 28.9, 28.2, 28.0, 27.9, 25.5, 22.6, 14.0; *anal.* C 71.57%, H 9.85%, calcd for C₃₆H₆₀O₇, C 71.49%, H 10.00%.

Bis-THF Alcohol 16. The procedure described for alcohol **10** was employed with 0.41 g (0.68 mmol) of olefin **15** and 0.41 g of Pd-C (5%) in 5.0 mL of EtOAc. The product was purified by column chromatography on Si gel (elution with 40% EtOAc in hexane) to afford 0.28 g (80%) of bis-THF alcohol **16**: IR (film) 3475 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 4.84 (1 H, d, J = 6.9 Hz), 4.82 (1 H, d, J = 6.9 Hz), 4.67 (2 H, d, J = 6.9 Hz), 4.07–3.83 (4 H, m), 3.64 (2 H, m), 3.48 (2 H, m), 3.39 (3 H, s), 3.37 (3 H, s), 2.00–1.16 (34 H, m), 0.87 (3 H, t, J = 7.2 Hz).

Bis-THF Aldehyde 17. To a mixture of 0.023 g (0.04 mmol) of alcohol 16 and 0.020 g of 4 Å molecular sieves in 1.0 mL of CH₂Cl₂ at 0 °C was added 0.020 g (0.09 mmol) of PCC. The reaction mixture was stirred at room temperature for 1 h, quenched with ether, and filtered through Celite. Solvent was removed under reduced pressure, and the residue was purified by column chromatography on Si gel (elution with 25% EtOAc in hexane) to afford 0.020 g (87%) of bis-THF aldehyde 17: $[\alpha]^{25}_{D}$ +42.8 (c 0.96, CHCl₃); IR (film) 1728 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 9.76 (1 H, s), 4.82 (1 H, d, J = 6.9 Hz), 4.81 (1 H, d, J = 6.9 Hz), 4.67 (1 H, d, J = 6.9 Hz), 4.65 (1 H, d, J = 6.9 Hz), 4.00 (2 H, m), 3.91 (2 H, m), 3.48 (2 H, m), 3.39 (3 H, s), 3.38 (3 H, s), 2.44 (2 H, dt, J = 1.5, 7.3 Hz), 2.00-1.16 (32 H, m), 0.87 (3 H, t, J = 7.2 Hz); ¹³C NMR (CDCl₃, 75 MHz) & 202.4, 96.7, 96.6, 81.7, 81.5, 81.3, 81.2, 81.1, 79.4, 79.2, 55.6, 43.7, 31.8, 31.1, 30.8, 29.7, 29.5, 29.2, 28.2, 25.5, 25.1, 22.6, 22.1, 14.0; anal. C 67.44%, H 10.79%, calcd for C₂₉H₅₄O₇, C 67.67% H 10.57%.

Hydroxy Ester 20. The previously described procedure^{4c} was employed with 0.66 g (0.13 mmol) of aldehyde **17** resulting in 0.045 g (50%) of alcohol **20**: $[\alpha]^{25}{}_{\rm D}$ +26.7 (*c* 0.38, CHCl₃); IR (film) 3501, 1736 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 4.82 (1 H, d, *J* = 6.9 Hz), 4.81 (1 H, d, *J* = 6.9 Hz), 4.67 (2 H, d, *J* = 6.9 Hz), 4.12 (2 H, q, *J* = 7.0 Hz), 4.00 (2 H, m), 3.91 (2 H, m), 3.57 (1 H, m), 3.47 (2 H, m), 3.39 (6 H, s), 2.28 (2 H, t, *J* = 7.3 Hz), 2.00–1.17 (51 H, m), 0.88 (3 H, t, *J* = 7.2 Hz); ¹³C NMR (CDCl₃, 75 MHz) δ 173.8, 96.6, 81.7, 81.6, 81.1, 79.4, 79.3, 71.7, 60.0, 55.6, 37.4, 37.3, 36.5, 34.3, 31.8, 31.1, 31.0, 30.0, 29.7, 29.5, 29.3, 29.2, 29.1, 29.0, 28.2, 25.7, 25.6, 25.5, 24.9, 22.6, 14.2, 14.0; *anal.* C 68.63%, H 10.94%, calcd for C₄₀H₇₆O₉, C 68.53%, H 10.93%.

(*R*)-Mandelate: ¹H NMR (CDCl₃, 300 MHz) δ 7.40 (5 H, m), 4.89 (1 H, m), 4.82 (1 H, d, J = 6.9 Hz), 4.78 (1 H, d, J = 6.9 Hz), 4.73 (1 H, s), 4.67 (1 H, d, J = 6.9 Hz), 4.61 (1 H, d, J = 6.9 Hz), 4.12 (2 H, q, J = 7.5 Hz), 3.94 (5 H, m), 3.47 (1 H, m), 3.41 (3 H, s), 3.39 (3 H, s), 3.36 (3 H, s), 2.28 (2 H, t, J = 7.7 Hz), 2.00–1.00 (49 H, m), 0.88 (3 H, t, J = 7.2 Hz).

(S)-Mandelate: ¹H NMR (CDCl₃, 300 MHz) δ 7.39 (5 H, m), 4.89 (1 H, m), 4.82 (1 H, d, J = 6.9 Hz), 4.82 (1 H, d, J = 6.9 Hz), 4.82 (1 H, d, J = 6.9 Hz), 4.81 (1 H, d, J = 6.9 Hz), 4.73 (1 H, s), 4.67 (1 H, d, J = 6.9 Hz), 4.65 (1 H, d, J = 6.9 Hz), 4.13 (2 H, q, J = 7.5 Hz), 3.99 (2 H, m), 3.91 (2 H, m), 3.46 (2 H, m), 3.41 (3 H, s), 3.39 (3 H, s), 3.38 (3 H, s), 2.27 (2 H, t, J = 7.7 Hz), 2.00–0.96 (49 H, m), 0.88 (3 H, t, J = 7.2 Hz).

MOM Ether 21. To a mixture of 0.036 g (0.05 mmol) of alcohol **20** and 0.10 mL (0.4 mmol) of *i*-Pr₂NEt in 0.50 mL of CH₂Cl₂ at 0 °C was added 0.02 mL (0.2 mmol) of MOMCl. The reaction mixture was stirred for 12 h, quenched with H₂O, and extracted with ether. The ether extracts were washed with brine, dried over MgSO₄, and concentrated under reduced pressure. The crude product was purified by column chromatography on Si gel (elution with 20% EtOAc in hexane) to afford 0.033 g (87%) of MOM ether **21**: $[\alpha]^{25}_D$ +33.5 (*c* 0.25, CHCl₃); IR (film) 1728 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 4.82 (2 H, d, *J* = 6.9 Hz), 4.66 (2 H, d, *J* = 6.9 Hz), 4.66 (2 H, m), 3.91 (2 H, m), 3.48 (3 H, m), 3.39 (6 H, s), 3.37 (3 H, s), 2.28 (2 H, t, *J* = 7.2 Hz), 2.00–1.15 (51 H, m), 0.88 (2 H, t, *J* = 7.2 Hz).

Lactone 23. To LDA prepared from 0.07 mL (0.18 mmol) of BuLi (2.5 M in hexane) and 0.03 mL (0.22 mmol) of *i*-Pr₂-

NH in 0.50 mL of THF at -78 °C was added a solution of 0.031 g (0.04 mmol) of ester 21 in 0.30 mL of THF. The reaction mixture was stirred at -78 °C for 1 h, and to it was added a solution of 0.016 g (0.08 mmol) of aldehyde 22 in 0.30 mL of THF. The reaction mixture was stirred at -78 °C for 1 h, quenched with NH₄Cl, and diluted with ether. The aqueous layer was extracted with ether, and the combined extracts were dried over MgSO₄. Solvent was removed under reduced pressure, and to the residue was added 1.0 mL of THF followed by 0.15 mL (0.15 mmol) of TBAF (1.0 M in THF). The reaction mixture was stirred for 1 h, quenched with H₂O, and extracted with ether. The extracts were dried over MgSO₄ and concentrated under reduced pressure. The crude product was purified by column chromatography on Si gel (elution with 40% EtOAc in hexane) to afford 0.022 g (69%) of lactone 23: IR (film) 3432, 1771 cm⁻¹; ¹H NMR (CDČl₃, 300 MHz) δ 4.82 (2 H, d, J = 6.9Hz), 4.66 (2 H, d, J = 6.9 Hz), 4.63 (2 H, s), 4.18 (1 H, m), 4.00 (2 H, m), 3.91 (2 H, m), 3.48 (3 H, m), 3.39 (6 H, s), 3.37 (3 H, s), 2.56 (1 H, m), 2.00–1.14 (51 H, m,), 0.88 (3 H, t, J = 7.2Hz).

Butenolide 25. To a mixture of 0.019 g (0.02 mmol) of alcohol **23** and 0.10 mL (0.2 mmol) of Et₃N in 2.0 mL of CH₂-Cl₂ at 0 °C was added 0.02 mL (0.08 mmol) of (CF₃CO)₂O. The reaction mixture was stirred at room temperature for 20 h, quenched with NaHCO₃, and extracted with ether. The extracts were dried over MgSO₄ and concentrated under reduced pressure. The crude product was purified by column chromatography on Si gel (elution with 20% EtOAc in hexane) to afford 0.017 g (90%) of butenolide **25**: IR (film) 1754 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 6.98 (1 H, s), 4.99 (1 H, m), 4.82 (2 H, d, *J* = 6.9 Hz), 4.66 (2 H, d, *J* = 6.9 Hz), 4.63 (2 H, s), 4.00 (2 H, m), 3.90 (2 H, m), 3.48 (3 H, m), 3.39 (6 H, s), 3.37 (3 H, s), 2.26 (2 H, t, *J* = 7.5 Hz), 2.00–1.12 (46 H, m), 1.40 (3 H, d, *J* = 6.9 Hz), 0.88 (3 H, t, *J* = 7.2 Hz).

Asimin (1). A mixture of 0.014 g (0.02 mmol) of butenolide 25 in 1.50 mL of 6 M HCl–THF–MeOH (1:2:2) was stirred for 12 h, quenched with H₂O, and extracted with ether. The ether extracts were dried over MgSO₄ and concentrated under reduced pressure. The crude product was purified by column chromatography on Si gel (elution with 80% of EtOAc in hexane) to afford 0.010 g (90%) of asimin (1): $[\alpha]^{25}_{D}$ +22.0 (*c* 0.45, CHCl₃), lit.⁵ +26.0 (*c* 0.10, CHCl₃); IR (film) 3459, 1745 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 6.98 (1 H, s), 4.99 (1 H, m), 3.86 (4 H, m), 3.58 (1 H, m), 3.39 (2 H, m), 2.26 (2 H, t, *J* = 7.6 Hz), 2.12–1.07 (46 H, m), 1.40 (3 H, d, *J* = 6.9 Hz), 0.87 (3 H, t, *J* = 7.3 Hz); ¹³C NMR (CDCl₃, 75 MHz) δ 173.8, 148.8, 134.3, 83.2, 83.0, 81.8, 74.1, 74.0, 71.8, 37.4, 33.4, 33.3, 31.9, 29.7, 29.6, 29.5, 29.3, 29.2, 29.1, 29.0, 28.0, 27.4, 25.7, 25.6, 25.1, 22.7, 19.2, 14.1.

Tri-(*S***)-Mosher Ester (26).** ¹H NMR (CDCl₃, 300 MHz) (diagnostic peaks are italicized) δ *5.00 (4 H, m)*, 3.94 (2 H, m), 3.78 (2 H, m), *3.54 (6 H, s)*, *3.52 (3 H, s)*, 2.26 (2 H, t, J = 7.7 Hz), 2.00–1.00 (46 H, m), 1.40 (3 H, d, J = 6.5 Hz), 0.88 (3 H, t, J = 6.5 Hz).

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Supporting Information Available: ¹H NMR spectra for all new compounds and experimental procedures for **17–34**. This information is available free of charge on the World Wide Web at http://pubs.acs.org.

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